

First-in-Human, First-in-Class Phase I Trial of the Anti-CD47 Antibody Hu5F9-G4 in Patients With Advanced Cancers

Branimir I. Sikic, MD¹; Nehal Lakhani, MD, PhD²; Amita Patnaik, MD³; Sumit A. Shah, MD¹; Sreenivasa R. Chandana, MD, PhD²; Drew Rasco, MD³; A. Dimitrios Colevas, MD¹; Timothy O'Rourke, MD²; Sujata Narayanan, MD¹; Kyriakos Papadopoulos, MD³; George A. Fisher, MD, PhD¹; Victor Villalobos, MD, PhD⁴; Susan S. Prohaska, PhD¹; Maureen Howard, PhD¹; Muralidhar Beeram, MD³; Mark P. Chao, MD, PhD⁵; Balaji Agoram, PhD⁵; James Y. Chen, MD, PhD⁵; Jie Huang, ScD⁵; Matthew Axt⁵; Jie Liu, PhD, MD⁵; Jens-Peter Volkmer, MD⁵; Ravindra Majeti, MD, PhD^{1,5}; Irving L. Weissman, MD¹; Chris H. Takimoto, MD, PhD⁵; Dana Supan, RN¹; Heather A. Wakelee, MD¹; Rhonda Aoki¹; Mark D. Pegram, MD¹; and Sukhmani K. Padda, MD¹

PURPOSE To evaluate the safety, pharmacokinetics, and pharmacodynamics of Hu5F9-G4 (5F9), a humanized IgG4 antibody that targets CD47 to enable phagocytosis.

PATIENTS AND METHODS Adult patients with solid tumors were treated in four cohorts: part A, to determine a priming dose; part B, to determine a weekly maintenance dose; part C, to study a loading dose in week 2; and a tumor biopsy cohort.

RESULTS Sixty-two patients were treated: 11 in part A, 14 in B, 22 in C, and 15 in the biopsy cohort. Part A used doses that ranged from 0.1 to 3 mg/kg. On the basis of tolerability and receptor occupancy studies that showed 100% CD47 saturation on RBCs, 1 mg/kg was selected as the priming dose. In subsequent groups, patients were treated with maintenance doses that ranged from 3 to 45 mg/kg, and most toxicities were mild to moderate. These included transient anemia (57% of patients), hemagglutination on peripheral blood smear (36%), fatigue (64%), headaches (50%), fever (45%), chills (45%), hyperbilirubinemia (34%), lymphopenia (34%), infusion-related reactions (34%), and arthralgias (18%). No maximum tolerated dose was reached with maintenance doses up to 45 mg/kg. At doses of 10 mg/kg or more, the CD47 antigen sink was saturated by 5F9, and a 5F9 half-life of approximately 13 days was observed. Strong antibody staining of tumor tissue was observed in a patient at 30 mg/kg. Two patients with ovarian/fallopian tube cancers had partial remissions for 5.2 and 9.2 months.

CONCLUSION 5F9 is well tolerated using a priming dose at 1 mg/kg on day 1 followed by maintenance doses of up to 45 mg/kg weekly.

J Clin Oncol 37. © 2019 by American Society of Clinical Oncology

INTRODUCTION

CD47 was first identified as an integrin-associated transmembrane protein that is ubiquitously found in normal and malignant tissues.¹ Most cancer cells overexpress CD47, and the degree of expression independently correlates with poor clinical outcome in a variety of hematologic and solid tumor malignancies.^{2,3} The binding of CD47 to its receptor signal receptor protein- α (SIRP α) on macrophages and dendritic cells results in an inhibition of phagocytosis. Thus, CD47 provides a potent “do not eat me” signal that allows for tumor cell evasion of immune destruction by first-responder phagocytic cells and functions as a dominant macrophage checkpoint.³⁻⁵ Blockade of CD47-SIRP α signaling in isolation is insufficient to trigger macrophage phagocytosis. Instead, additional pro-phagocytic signals are required, such as calreticulin

and phosphatidylserine, which are frequently found on cancer cells.^{6,7} CD47 also is widely expressed on normal tissues, but because normal cells lack pro-phagocytic signals, they are not susceptible to CD47-mediated phagocytosis. A notable exception is aging RBCs.⁸ Agents that inhibit CD47-SIRP α signaling can induce macrophage phagocytosis of cancer cells both in vitro and in vivo, which results in growth inhibition and regression of a broad range of human cancer xenografts.^{3,4} Therefore, the targeting of CD47 is a novel immunotherapeutic strategy for treating human cancers.

Hu5F9-G4 (5F9) is a humanized IgG4 monoclonal antibody with high affinity for human CD47.⁹ 5F9-mediated blockade of CD47 enhances the phagocytosis of cancer cells by macrophages. In preclinical in vivo models, 5F9 was active against a wide range of

ASSOCIATED CONTENT

Appendix

Data Supplements

See accompanying article doi: [10.1200/JCO.19.00121](https://doi.org/10.1200/JCO.19.00121)

Author affiliations and support information (if applicable) appear at the end of this article.

Accepted on January 30, 2019 and published at [jco.org](https://doi.org/10.1200/JCO.18.02018) on February 27, 2019; DOI <https://doi.org/10.1200/JCO.18.02018>

Processed as a rapid communication manuscript.

Clinical trial information: NCT02216409.

solid tumors, including cancers of the breast, ovary, colon, liver, brain, and other organs.³⁻⁵ Potent antitumor activity also was observed in hematologic malignancies, including acute myeloid leukemia (AML), non-Hodgkin lymphoma, cutaneous T-cell lymphoma, acute lymphoblastic leukemia, and multiple myeloma.⁹ In human tumor xenograft models, 5F9 inhibited tumor cell growth and induced remission in established tumors.⁹ In preclinical toxicology studies, the major dose-limiting toxicity (DLT) was an on-target anemia¹⁰ that was mitigated by using a priming and maintenance dose schedule. Using this approach, non-human primates tolerated 5F9 doses up to 300 mg/kg without reaching a maximum tolerated dose (MTD).⁹

This report describes the first-in-human phase I trial of 5F9 in patients with advanced solid tumors and lymphomas. The trial consisted of three distinct dose escalation parts. Part A used weekly dosing to determine a tolerable day 1 priming dose. Part B administered the 5F9 priming dose identified in part A followed by escalation of weekly maintenance doses to establish an MTD. At the completion of part B, a tumor biopsy expansion cohort was opened. In part C, a loading dose was given on day 11 in addition to weekly 5F9 therapy to enable more-rapid attainment of therapeutic concentrations. The safety, tolerability, and early efficacy results along with summary pharmacokinetics (PK) and pharmacodynamics (PD) data are described here. Detailed PK and PD findings will be reported elsewhere.

PATIENTS AND METHODS

Patient Selection and Oversight

Eligible patients were adults 18 years of age or older with histologically or cytologically confirmed advanced solid malignancy or lymphoma previously treated with at least one regimen of systemic therapy, or who refused other systemic therapy, with an Eastern Cooperative Oncology Group performance status of 0 to 2 and for whom no curative therapy was available. Additional eligibility criteria are outlined in the Data Supplement.

This study was approved by the human investigations committee at each institution, and in accordance with assurances filed with and approved by the Department of Health and Human Services. These data are anonymized to protect the identities of the research participants. Informed written consent was obtained from each participant. Patients were enrolled from August 2014 through January 2018.

Study Design and Drug Administration

The primary objectives were to assess the safety and tolerability of 5F9 and to determine the recommended doses and schedules for additional trials. The secondary objectives were to evaluate the PK and PD profiles and to document antitumor activity. Treatment cycles were 28 days long and continued until disease progression. In

part A, the 5F9 starting dose of 0.1 mg/kg was infused intravenously weekly, with dose escalation conducted in single-patient cohorts for doses less than 3 mg/kg in a modified accelerated titration design.¹¹ The duration of the infusion was 1 hour for doses from 0.1 to 1 mg/kg and 2 hours for 3 mg/kg. In part B and beyond, all patients received a 1 mg/kg priming dose on day 1 for 3 hours followed by escalating maintenance doses for 2 hours on day 8 that began at the 3 mg/kg weekly dose level. After a safe dose was determined in part B, a solid tumor expansion cohort of 15 patients was opened, which required mandatory pretreatment and during treatment tumor biopsies.

Beginning in part B, routine premedication with acetaminophen and an antihistamine was administered before the first two doses of 5F9. In part C, patients received a loading dose on day 11 of cycle 1 (cohorts of 20, 30, and 45 mg/kg) in addition to weekly maintenance dosing at the same level. After cycle 1, if patients had treatment delays longer than 4 weeks, repriming was required.

In part B, dose escalation was performed using a traditional 3 + 3 phase I study design.¹² DLT was defined as any drug-related adverse event (AE) of grade 3 or greater that occurred from the time of first dose in part A, or from the first maintenance doses in parts B and C, until the completion of 28 days of therapy (cycle 1), with exceptions that are listed in the Appendix. If a DLT was observed, the number of patients at that dose level was expanded from three up to six. Grade 3 anemia was excluded as a DLT because it was expected and anticipated to be transient and reversible. The MTD was defined as the highest dose level explored that induced DLTs in less than 33% of patients. Up to 10 more patients could be entered to any safe dose level to collect additional safety and PK data.

Study Assessments

All patients who received at least one 5F9 treatment were evaluated for safety. Standard safety assessments were conducted at baseline, before each weekly infusion, and out to 28 days after the last dose of study drug. Because of the possibility of hemagglutination, peripheral blood smear, visual acuity, and retinal photographic examinations were performed three times weekly for the first two weeks in part A. In part B, peripheral blood smears were examined on days 1, 2, 8, 9, 15, 22, and 29, and visual acuity was measured on days 1 and 8. However, there were no adverse findings on retinal photography as assessed by an ophthalmologist in part A. Therefore, retinal photography was not performed in parts B and C. Routine laboratory assessments are detailed in the Appendix. AEs were graded according to the National Cancer Institute Common Terminology Criteria for Adverse Events (version 4.03). Tumor measurements were obtained at baseline and every 8 weeks until disease progression or study withdrawal. Response was determined using Response Evaluation Criteria in Solid Tumors (RECIST) version 1.1¹³ or the International Working Group response criteria for

malignant lymphomas.¹⁴ Details on PK and antidrug antibody (ADA) monitoring as well as PD receptor occupancy studies and tumor biopsy analyses are provided in the Data Supplement.

Statistical Analysis

This was an open-label, nonrandomized, first-in-human dose escalation phase I trial. All patients exposed to 5F9 were included in the safety analysis, and all patients who had at least one postbaseline tumor assessment were considered evaluable for tumor response assessment.

RESULTS

Patient Characteristics

The characteristics of all 62 treated patients are listed in Table 1. Female patients comprised 65% of participants

and included 13 with ovarian cancer and five with breast cancer. The most frequent tumor type was colorectal cancer. Nine patients had salivary gland or head and neck cancers, and two had diffuse large B-cell lymphoma (DLBCL). Patients were heavily pretreated, having received a median of five systemic treatments (range, one to 18 treatments) before study enrollment. The Eastern Cooperative Oncology Group performance status was 0 in 19 patients (30%), 1 in 42 patients (68%), and 2 in one patient (2%).

Safety

The treatment cohorts and DLTs are listed in Table 2. Two DLTs were observed in part A at 3 mg/kg, including one patient who developed grade 3 right-side upper-abdominal pain during the first infusion of 5F9 and one patient with grade 3 hemagglutination defined as grade 2 headache associated with hemagglutination on the peripheral blood smear. However, no DLTs were observed in the six patients treated at 1 mg/kg, so this dose was selected as the day 1 priming dose for use in all subsequent parts of the study.

In part B, weekly maintenance doses were administered at 3, 10, and 20 mg/kg beginning on day 8 after the priming dose. One patient at 3 mg/kg had transient grade 3 elevated bilirubin without changes in serum transaminases, which at the time was defined as a DLT. A second patient treated at 20 mg/kg had grade 3 hemagglutination (grade 2 headache with hemagglutination on peripheral blood smear), but no other DLTs occurred in the six patients treated at this dose level. Fifteen additional patients were treated with 20 mg/kg in the tumor biopsy expansion cohort to collect additional PD data, and no DLTs were observed. In part C, one patient at 20 mg/kg experienced grade 3 elevated bilirubin, but no additional DLTs occurred despite escalation to 30 mg/kg and 45 mg/kg in nine and six patients, respectively. Thus, no MTD was reached in parts B or C. On the basis of this favorable safety profile and the accumulated PK data that demonstrated steady-state dose levels well above the targeted threshold of 200 µg/mL¹⁵ (Fig 1A), a maintenance dose of 30 mg/kg was recommended for additional clinical evaluation.

The combined AE profile from 44 patients treated with 5F9 maintenance doses of 20, 30, and 45 mg/kg are listed in Table 3. Overall, the most common 5F9 toxicity was the expected on-target anemia, which occurred in 57% of patients treated with doses that ranged from 20 to 45 mg/kg (13% of patients with grade 3). Typically, a modest, predictable fall in hemoglobin (1 to 2 g/dL) occurred after the 1 mg/kg priming dose followed by a rapid compensatory reticulocytosis, as illustrated in a single patient in Figure 1B. 5F9 treatment was not associated with thrombocytopenia. In the 52 patients treated for at least 4 weeks, the median percent change in platelet count from baseline on day 29 at the end of the first cycle was -3.2% (range, -54.3% to 91.2%). Five patients received concomitant anticoagulant therapy, but no associated bleeding or thrombocytopenia was observed.

TABLE 1. Patient Demographics and Baseline Characteristics

Characteristic	No.	%
No. of patients treated	62	100
Median age, years (range)	60 (35-84)	
Sex		
Male	22	35
Female	40	65
Ethnicity		
Hispanic or Latino	4	6.5
Not Hispanic or Latino	58	93.5
Race		
White	49	79.0
Asian	11	17.7
Black	2	3.2
ECOG performance status		
0	19	30
1	42	68
2	1	2
Median No. of prior systemic treatments (range)	5 (1-18)	
Tumor type		
Colorectal	18	29.0
Ovarian	13	21.0
Salivary	5	8.1
Breast	5	8.1
Head and neck	4	6.5
Pancreatic	4	6.5
Lung	2	3.2
Skin	2	3.2
DLBCL	2	3.2
Other	7	11.3

Abbreviations: DLBCL, diffuse large B-cell lymphoma; ECOG, Eastern Cooperative Oncology Group.

TABLE 2. Treatment Cohorts and DLTs

Part and Dose Cohort	Patients, No.	Infusions, No.	Patients With DLTs, No.
Part A: weekly dose (prime), mg/kg			
0.1	1	72	0
0.3	2	11	0
1	6	70	0
3	2	2	2*†
Part B: prime, maintenance doses, mg/kg			
1, 3	4	20	1‡
1, 10	3	112	0
1, 20	7	78	1†
Tumor biopsy cohort: prime, maintenance doses, mg/kg			
1, 20	15	197	0
Part C: prime, loading, maintenance doses, mg/kg			
1, 20, 20	7	130	1‡
1, 30, 30	9	129	0
1, 45, 45	6	54	0
Total	62	875	5

Abbreviation: DLT, dose-limiting toxicity.

*Grade 3 pain, right-side upper abdomen.

†Grade 3 hemagglutination (grade 2 headache with hemagglutination on peripheral blood smear).

‡Grade 3 elevated bilirubin (unconjugated)/blood bilirubin increase.

Hemagglutination on the peripheral blood smear was noted in 36% of patients but was not associated with significant clinical toxicities (2% grade 3). Specifically, no neurologic, renal, or cutaneous abnormalities were noted.

Other common toxicities were grade 1 and 2 fatigue (64%), headaches (50%; 2% grade 3), fever (45%), chills (45%), hyperbilirubinemia (34%), and arthralgias (18%). Unconjugated hyperbilirubinemia that was not associated with other liver function test abnormalities was common after the 5F9 priming dose; however, it generally returned to baseline within a few days. Transient elevation in unconjugated bilirubin was likely due to the expected phagocytosis of aged RBCs that result from blockade of CD47. As such, in part B, the protocol was amended to exclude grade 3 hyperbilirubinemia from the DLT definition. First- or second-dose infusion-related reactions (IRRs) occurred in 34% of patients, predominantly in those enrolled in the 20 mg/kg and higher dose cohorts. These were characterized by symptoms such as chills, fever/pyrexia, photopsia, back pain, and headaches that generally lasted a day or two after the associated infusion and were managed with symptomatic supportive care. Overall, only one patient underwent dose reduction from 30 to 20 mg/kg in

part C because of an IRR, and this patient was able to receive further treatment without problems. One additional patient withdrew consent because of infusion-related AEs. However, all other patients who experienced IRRs were able to continue treatment at their assigned dose level.

Transient grade 3 IRRs that responded to symptomatic treatment and that did not require dose reductions were not considered DLTs. No risk factors for IRRs were identified. After the first three patients were treated in the part C loading dose cohort at 30 mg/kg, routine antihistamine and acetaminophen premedication was mandated for all new patients before the initial priming dose of 1 mg/kg on day 1 and before the second higher dose on day 8. However, even with routine prophylaxis, grade 1 or 2 IRRs that were managed symptomatically were common and reported in seven of 12 premedicated patients treated in part C at 30 or 45 mg/kg. Lymphopenia was found in 36% of patients without apparent clinical consequences. No treatment-related deaths occurred on study.

Hemagglutination was seen on peripheral blood smear typically after the first or second 5F9 dose. However, there were no clear clinical sequelae. Specifically, no treatment-related peripheral vascular symptoms, changes in renal function, deep venous thromboses, pulmonary emboli, or other thrombotic events were observed. Furthermore, serial retinal photographs were collected over seven time points in the first cycle to assess for microvascular changes, but no such findings were noted, so the protocol was amended to remove retinal photography evaluation in part B.

PK and ADA Analysis

Similar to the PK in nonhuman primates, 5F9 exhibited nonlinear PK behavior as a result of the CD47 antigen sink, which resulted in a short apparent half-life at doses of 0.1 to 3 mg/kg of 5F9 (Fig 1A; and online only Appendix Table A1). However, doses of 10 mg/kg and higher saturated this sink, which extended the 5F9 half-life to approximately 13 days, consistent with other humanized monoclonal antibodies.¹⁶ At these doses, free antibody levels exceeded the antitumor activity threshold of 200 µg/mL identified from PK modeling.^{9,15,19} Serum samples from six (9.7%) of 62 patients tested positive for ADA, but there was no impact on 5F9 PK and no apparent clinical symptoms.¹⁵ Detailed PK profiling will be presented elsewhere.

PD and Tissue Biopsy Analysis

Studies of RBC occupancy showed complete saturation of CD47 on RBCs at doses of 1 mg/kg and higher (Fig 2A), which also supported 1 mg/kg as the initial 5F9 priming dose. In contrast, maximal saturation of CD47 on WBCs was only observed after higher maintenance doses (Fig 2B). Doses of 30 mg/kg maximally saturated CD47 on circulating WBCs.

In addition, anti-IgG4 staining demonstrated strong antibody penetration into metastatic tumor tissue in an axillary

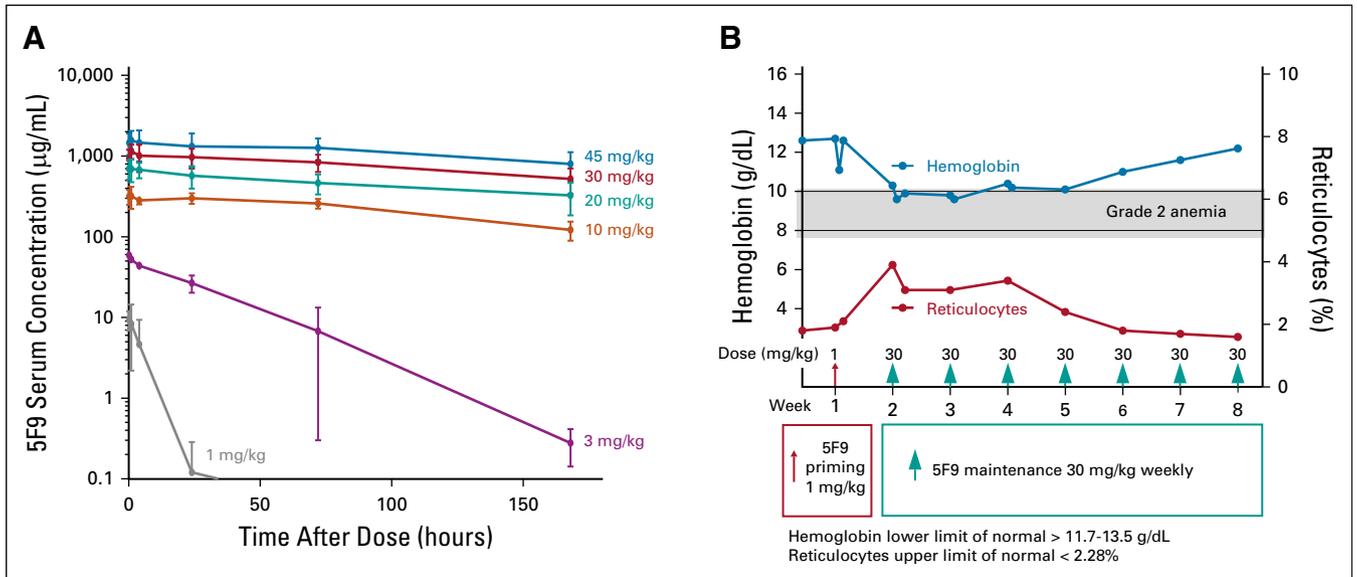


FIG 1. Hu5F9-G4 (5F9) pharmacokinetics and effects on hemoglobin and reticulocyte levels. (A) Concentration of 5F9 (mean ± standard deviation) versus time during week 5 over the evaluated weekly dose range (1 to 45 mg/kg; number of patients per dose range, three to 30). (B) Anemia with compensatory reticulocytosis in one patient.

lymph node from a patient with metastatic ovarian cancer (Fig 2C). This patient was treated with 30 mg/kg 5F9, and the biopsy sample was obtained after 5 weeks of treatment. These collected data support the recommended dose of 30 mg/kg for phase II evaluation.

Antitumor Activity

Two patients with clear cell ovarian and fallopian tube carcinomas had confirmed partial responses by RECIST

version 1.1 (Figs 3A and 3B), with time to progression of 5.2 months and 9.2 months, respectively. These patients had reductions in target lesions of 50% and 44%. Their cancer antigen 125 tumor markers dropped from 338 to 70 U/mL and 890 to 103 U/mL, respectively. Both patients were heavily pretreated with more than six prior lines of systemic therapy, and both received 5F9 at 20 mg/kg. A third patient with DLBCL at the 20 mg/kg maintenance dose had a mixed response by International Working Group

TABLE 3. Adverse Event Summary for Patients Treated With Maintenance Doses of 20 mg/kg or Higher
Patients Treated, No. (%)

Adverse Event*	Part B, Biopsy Cohort, and Part C Prime + 20 mg/kg Maintenance Dose (n = 29)			Part C: Prime + 30 mg/kg Load and Maintenance Dose (n = 9)			Part C: Prime + 45 mg/kg Load and Maintenance Dose (n = 6)		
	All Grades	Grade 3	Grade 4	All Grades	Grade 3	Grade 4	All Grades	Grade 3	Grade 4
Anemia	19 (66)	5 (17)	0	3 (33)	0	0	3 (50)	1 (17)	0
Hemagglutination	12 (41)	1 (3)	0	2 (22)	0	0	2 (33)	0	0
Hyperbilirubinemia	11 (38)	3 (10)	0	0	0	0	1 (17)	0	0
Thrombocytopenia	5 (17)	0	0	0	0	0	0	0	0
Lymphocyte count decreased	4 (14)	4 (14)	0	3 (33)	2 (22)	1 (11)	1 (17)	1 (17)	0
Arthralgia/myalgia	5 (17)	0	0	2 (22)	0	0	1 (17)	0	0
Headache	11 (38)	0	0	6 (67)	1 (11)	0	4 (67)	0	0
Nausea	3 (10)	0	0	2 (22)	0	0	3 (50)	0	0
Fatigue	18 (62)	0	0	6 (67)	0	0	4 (67)	0	0
Fever	14 (48)	0	0	4 (44)	0	0	2 (33)	0	0
Chills	12 (41)	0	0	5 (56)	0	0	3 (50)	0	0
Infusion-related reaction	7 (24)	2 (7)	0	2 (22)	1 (11)	0	2 (33)	1 (17)	0

*Adverse events occurred in > 15% of patients across all three cohorts listed (n = 44) and selected adverse events of interest.

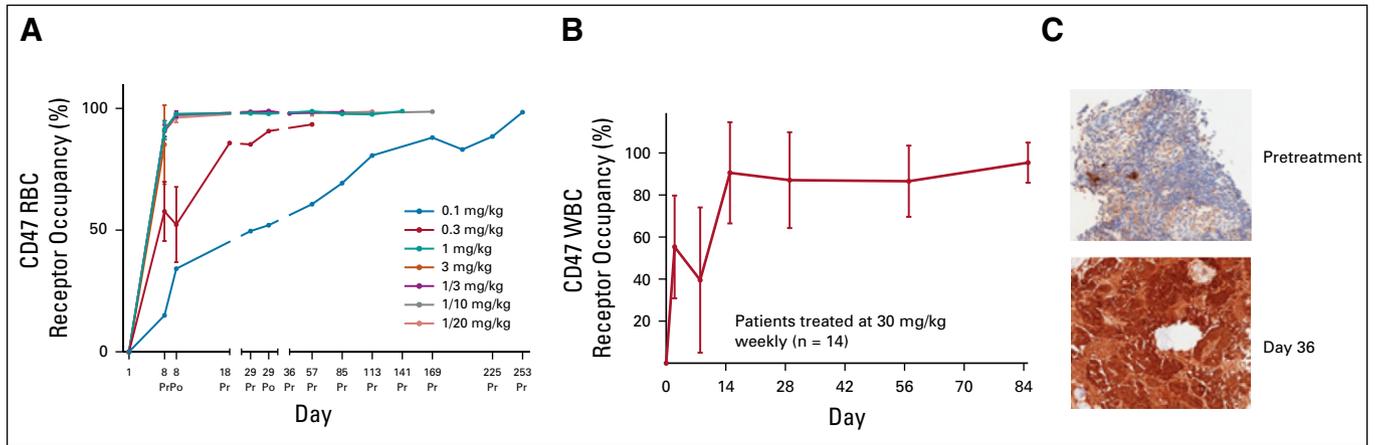


FIG 2. Pharmacodynamics of Hu5F9-G4. (A) RBC receptor occupancy. (B) WBC receptor occupancy. (C) IgG4 immunohistochemical staining in a lymph node with tumor involvement obtained from a 70-year-old patient with ovarian cancer treated with 30 mg/kg of 5F9. She remained on study for 7 months with a best overall response of stable disease. 1/3 is 1 mg/kg priming dose and 3 mg/kg weekly dosing. 1/10 is 1 mg/kg priming dose and 10 mg/kg weekly dosing. 1/20 is 1 mg/kg priming dose and 20 mg/kg weekly dosing. Po, post-infusion sample collection; Pr, pre-infusion sample collection.

criteria. This heavily pretreated patient with non-Hodgkin lymphoma had bulky lesions in the chest that decreased in size but had a solitary inguinal lymph node that increased while on treatment.

DISCUSSION

The anti-CD47 antibody 5F9 is well-tolerated in patients with solid tumors and lymphoma when administered using a priming and maintenance dose regimen. An MTD was not defined, even at doses up to 45 mg/kg weekly. The recommended 5F9 priming dose of 1 mg/kg on day 1 achieved maximal CD47 receptor occupancy on RBCs. Receptor occupancy on circulating WBCs was not available at low maintenance doses, but available data confirmed complete WBC receptor occupancy at weekly maintenance dosing of 30 mg/kg beginning on day 8. A tissue biopsy sample of a lymph node with metastatic ovarian cancer in one patient demonstrated excellent penetration of 5F9 into the tumor microenvironment. At doses above 10 mg/kg, the PK of 5F9 was linear, and increases in drug exposure were dose proportional. The resulting long terminal half-life of approximately

13 days supports every 2-week dosing after saturating concentrations are achieved.

The most common toxicity was expected, on-target, mild, transient, and predictable anemia, which was mitigated by administering priming and maintenance doses in cycle 1. After the initial priming dose, hemoglobin levels dropped on average by approximately 2 g/dL presumably because of RBC phagocytosis by macrophages in the liver and spleen. However, this was immediately followed by a rapid compensatory reticulocytosis as younger RBCs were released into circulation. Younger RBCs are unperturbed by CD47 blockade because they lack pro-phagocytic signals that accumulate on older RBCs. Despite continued 5F9 dosing, hemoglobin levels typically remained stable beyond the first week, and in many patients, the anemia gradually resolved to baseline as the circulating RBC population shifted to a younger average age. Hemagglutination on peripheral blood smear was noted in some patients, typically after the first or second dose, but there was no consistent association with clinical sequelae. Blood transfusions were infrequently required in this mostly solid tumor population, with four of

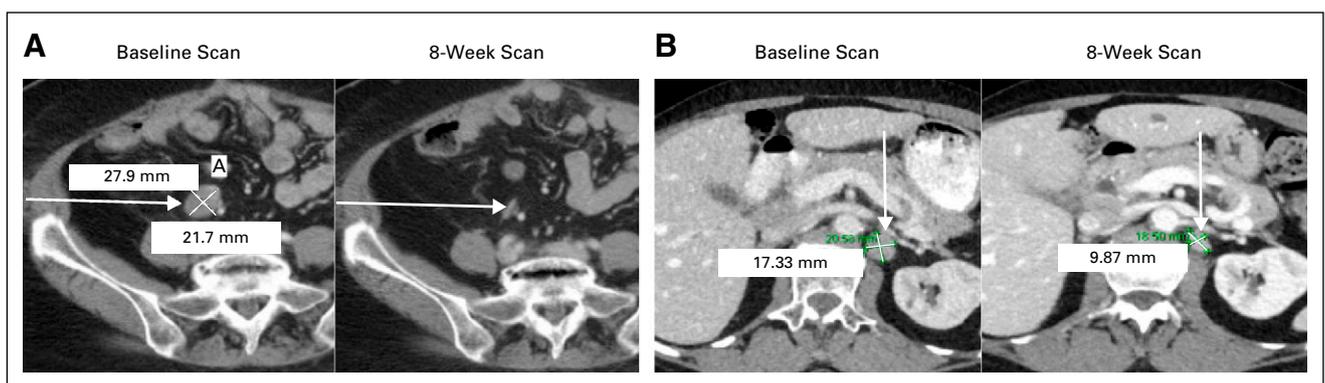


FIG 3. Hu5F9-G4 partial responses observed in (A) a patient with clear cell ovarian cancer and (B) a patient with fallopian tube cancer.

62 patients receiving RBC transfusions. However, early clinical experience with 5F9 in patients with refractory end-stage AML¹⁷ suggests that blood product support can readily be administered to patients who receive 5F9 therapy.

Most 5F9-related AEs were mild to moderate in severity, and 5F9 infusions were easily administered in an outpatient setting. At dose levels of 20 mg/kg and higher, the commonly observed AEs were headache, fatigue, fever, and chills during the infusions, with most occurring within the first cycle. Pretreatment with acetaminophen and diphenhydramine lowered the incidence and severity of these reactions, and this pretreatment is recommended before the first two doses of 5F9. A few patients experienced abdominal, back, or chest pains during infusion that were not associated with other clinical findings and often was mitigated by temporarily stopping or slowing the rate of infusion. No treatment-related autoimmune phenomena (pneumonitis, colitis, or hepatitis) analogous to those reported with programmed cell death 1/programmed cell death-ligand 1 checkpoint inhibitors were observed. Despite the widespread expression of CD47 on normal tissues, patients tolerated 5F9 well presumably because normal cells, unlike their malignant counterparts, lack phagocytic signals and thus are not susceptible to CD47 blockade. Of note, no dose-related thrombocytopenia or neutropenia was observed. This favorable safety profile suggests that 5F9 may be readily combined with other antitumor antibodies, which is consistent with emerging clinical reports.¹⁸

Of note, single-agent 5F9 induced objective tumor responses in two patients, one with clear cell ovarian cancer and one with a fallopian tube cancer. Both patients were

heavily pretreated with more than six lines of systemic therapy, including multiple lines of platinum-based chemotherapy. However, all phase I first-in-human studies have limitations because of the heterogeneity of patients who may not reflect broader phase II and III study populations because they are selected for more indolent disease, including many who have participated in multiple prior phase I studies. Therefore, further evaluation of 5F9 is ongoing in an additional cohort of patients with ovarian cancer.

Detailed translational studies that analyze treatment-related changes in the tumor microenvironment in biopsy samples from patients in this trial are ongoing. Additional trials are ongoing that are evaluating 5F9 therapy in patients with ovarian cancer, AML, and cutaneous T-cell lymphoma. 5F9 generated objective responses in patients with solid tumors in this phase I trial; however, its optimal therapeutic impact likely will be in combination with other anticancer therapies.¹⁸ Ongoing combination trials are evaluating 5F9 plus rituximab in B-cell lymphomas (ClinicalTrials.gov identifier: NCT02953509),¹⁸ with cetuximab in colorectal cancers (ClinicalTrials.gov identifier: NCT02953782), with azacitidine in treatment-naïve AML and myelodysplastic syndrome (ClinicalTrials.gov identifier: NCT03248479), and with the programmed cell death-ligand 1 checkpoint inhibitor avelumab in patients with ovarian cancer (ClinicalTrials.gov identifier: NCT03558139). 5F9 has received fast-track designation by the US Food and Drug Administration for further development in DLBCL and follicular lymphoma. In summary, enhancement of macrophage phagocytosis of tumor cells by blocking CD47 with 5F9 is a promising new cancer therapeutic strategy.

AFFILIATIONS

¹Stanford University School of Medicine, Stanford, CA

²South Texas Accelerated Therapeutics Midwest, Grand Rapids, MI

³South Texas Accelerated Therapeutics, San Antonio, TX

⁴University of Colorado Cancer Center, Aurora, CO

⁵Forty Seven, Menlo Park, CA

CORRESPONDING AUTHOR

Branimir I. Sikic, MD, Division of Oncology, Department of Medicine, Stanford University School of Medicine, 269 Campus Dr, CCSR-1105c, Stanford, CA 94305; e-mail: brandy@stanford.edu.

PRIOR PRESENTATION

Presented at the American Society of Clinical Oncology 2018 Annual Meeting, Chicago, IL, June 1-5, 2018.

SUPPORT

Supported by grants from the National Institutes of Health (Stanford Clinical and Translational Research Unit), California Institute for Regenerative Medicine, and Forty Seven.

AUTHORS' DISCLOSURES OF POTENTIAL CONFLICTS OF INTEREST AND DATA AVAILABILITY STATEMENT

Disclosures provided by the authors and data availability statement (if applicable) are available with this article at DOI <https://doi.org/10.1200/JCO.18.02018>.

AUTHOR CONTRIBUTIONS

Conception and design: Branimir I. Sikic, Sumit A. Shah, A. Dimitrios Colevas, Sujata Narayanan, George A. Fisher, Victor Villalobos, Susan S. Prohaska, Maureen Howard, Mark P. Chao, Jie Huang, Jie Liu, Jens-Peter Volkmer, Ravindra Majeti, Irving L. Weissman, Chris H. Takimoto, Mark D. Pegram, Sukhmani K. Padda

Financial support: Branimir I. Sikic, Ravindra Majeti, Chris H. Takimoto

Administrative support: Branimir I. Sikic, Nehal Lakhani, Sreenivasa R. Chandana, Ravindra Majeti, Chris H. Takimoto, Rhonda Aoki

Provision of study material or patients: Branimir I. Sikic, Nehal Lakhani, Drew Rasco, A. Dimitrios Colevas, Timothy O'Rourke, Kyriakos Papadopoulos, George A. Fisher, Susan S. Prohaska, Heather A. Wakelee, Mark D. Pegram

Collection and assembly of data: Branimir I. Sikic, Nehal Lakhani, Amita Patnaik, Sumit A. Shah, Sreenivasa R. Chandana, Drew Rasco, A. Dimitrios Colevas, Timothy O'Rourke, Sujata Narayanan, Kyriakos Papadopoulos, George A. Fisher, Victor Villalobos, Maureen Howard, Muralidhar Beeram, Mark P. Chao, Balaji Agoram, James Y. Chen, Jie

Huang, Matthew Axt, Jens-Peter Volkmer, Chris H. Takimoto, Dana Supan, Heather A. Wakelee, Rhonda Aoki, Mark D. Pegram, Sukhmani K. Padda

Data analysis and interpretation: Branimir I. Sikic, Nehal Lakhani, Amita Patnaik, Sumit A. Shah, A. Dimitrios Colevas, Sujata Narayanan, George A. Fisher, Maureen Howard, Mark P. Chao, Balaji Agoram, James Y. Chen, Jie Huang, Jens-Peter Volkmer, Irving L. Weissman, Chris H. Takimoto, Heather A. Wakelee, Mark D. Pegram, Sukhmani K. Padda
Manuscript writing: All authors

Final approval of manuscript: All authors

Accountable for all aspects of the work: All authors

ACKNOWLEDGMENT

We thank Susan Jerian, MD, and David Essayan, MD, of ONCORD for their advice on study design. We are most grateful to the participating patients and their family members and to all the nurses, investigators, and study staff who contributed to this clinical study.

REFERENCES

1. Brown EJ, Frazier WA: Integrin-associated protein (CD47) and its ligands. *Trends Cell Biol* 11:130-135, 2001
2. Chao MP, Alizadeh AA, Tang C, et al: Anti-CD47 antibody synergizes with rituximab to promote phagocytosis and eradicate non-Hodgkin lymphoma. *Cell* 142:699-713, 2010
3. Willingham SB, Volkmer JP, Gentles AJ, et al: The CD47-signal regulatory protein alpha (SIRPα) interaction is a therapeutic target for human solid tumors. *Proc Natl Acad Sci U S A* 109:6662-6667, 2012
4. Chao MP, McKenna KM, Cha A, et al: The anti-CD47 antibody Hu5F9-G4 is a novel immune checkpoint inhibitor with synergistic efficacy in combination with clinically active cancer targeting antibodies. *Cancer Immunol Res* 4, 2016 (abstr PR13)
5. Majeti R, Chao MP, Alizadeh AA, et al: CD47 is an adverse prognostic factor and therapeutic antibody target on human acute myeloid leukemia stem cells. *Cell* 138:286-299, 2009
6. Chen Q, Fang X, Jiang C, et al: Thrombospondin promoted anti-tumor of adenovirus-mediated calreticulin in breast cancer: Relationship with anti-CD47. *Biomed Pharmacother* 73:109-115, 2015
7. Chao MP, Jaiswal S, Weissman-Tsukamoto R, et al: Calreticulin is the dominant pro-phagocytic signal on multiple human cancers and is counterbalanced by CD47. *Sci Transl Med* 2:63ra94, 2010
8. Oldenburg PA, Zheleznyak A, Fang YF, et al: Role of CD47 as a marker of self on red blood cells. *Science* 288:2051-2054, 2000
9. Liu J, Wang L, Zhao F, et al: Pre-clinical development of a humanized anti-CD47 antibody with anti-cancer therapeutic potential. *PLoS One* 10:e0137345, 2015
10. Olsson M, Oldenburg PA: CD47 on experimentally senescent murine RBCs inhibits phagocytosis following Fcγ receptor-mediated but not scavenger receptor-mediated recognition by macrophages. *Blood* 112:4259-4267, 2008
11. Simon R, Freidlin B, Rubinstein L, et al: Accelerated titration designs for phase I clinical trials in oncology. *J Natl Cancer Inst* 89:1138-1147, 1997
12. Le Tourneau C, Lee JJ, Siu LL: Dose escalation methods in phase I cancer clinical trials. *J Natl Cancer Inst* 101:708-720, 2009
13. Eisenhauer EA, Therasse P, Bogaerts J, et al: New response evaluation criteria in solid tumours: Revised RECIST guideline (version 1.1). *Eur J Cancer* 45:228-247, 2009
14. Barrington SF, Mikhaeel NG, Kostakoglu L, et al: Role of imaging in the staging and response assessment of lymphoma: Consensus of the International Conference on Malignant Lymphomas Imaging Working Group. *J Clin Oncol* 32:3048-3058, 2014
15. Agoram B, Wang B, Sikic BI, et al: Pharmacokinetics of Hu5F9-G4, a first-in-class anti-CD47 antibody, in patients with solid tumors and lymphomas. *J Clin Oncol* 36, 2018 (suppl; abstr 2525)
16. Wang W, Wang EQ, Balthasar JP: Monoclonal antibody pharmacokinetics and pharmacodynamics. *Clin Pharmacol Ther* 84:548-558, 2008
17. Vyas P, Knapper S, Kelly R, et al: Initial phase I results of the first-in-class anti-CD47 antibody Hu5F9-G4 in relapsed/refractory acute myeloid leukemia patients. *EHA Learning Center* 2018 (abstr PF232)
18. Advani R, Flinn I, Popplewell L, et al: CD47 blockade by Hu5F9-G4 and rituximab in non-Hodgkin's lymphoma. *N Engl J Med* 379:1711-1721, 2018
19. Buckeridge C, Duwuri S, Denney WS: Simple, automatic noncompartmental analysis: The PKNCA R package. *J Pharmacokinet Pharmacodyn* 42:S65, 2015



AUTHORS' DISCLOSURES OF POTENTIAL CONFLICTS OF INTEREST

First-in-Human, First-in-Class Phase I Trial of the Anti-CD47 Antibody Hu5F9-G4 in Patients With Advanced Cancers

The following represents disclosure information provided by authors of this manuscript. All relationships are considered compensated. Relationships are self-held unless noted. I = Immediate Family Member, Inst = My Institution. Relationships may not relate to the subject matter of this manuscript. For more information about ASCO's conflict of interest policy, please refer to www.asco.org/rwc or ascopubs.org/jco/site/ffc.

Branimir I. Sikic

Consulting or Advisory Role: Threshold Pharmaceuticals, Immune Design
Research Funding: Forty Seven (Inst), Celldex Therapeutics (Inst), Gilead Sciences (Inst), Basilea Pharmaceutica (Inst), Genentech (Inst), Roche (Inst), Sanofi (Inst)

Nehal Lakhani

Research Funding: Alexo Therapeutics (Inst), Ascentage Pharma (Inst), Asana BioSciences (Inst), BeiGene (Inst), Constellation Pharmaceuticals (Inst), Alexion Pharmaceuticals (Inst), Cerulean Pharma (Inst), Forty Seven (Inst), Loxo Oncology (Inst), Macrogenics (Inst), Merck (Inst), Pfizer (Inst), Regeneron Pharmaceuticals (Inst), TaiRx (Inst), Apexian Pharmaceuticals (Inst), Formation Biologics (Inst), Coordination Therapeutics (Inst), Symphogen (Inst), CytomX Therapeutics (Inst), InhibRx (Inst), Incyte (Inst), Jounce Therapeutics (Inst), Livzon Pharmaceutical Group (Inst), Northern Biologics (Inst), TESARO (Inst)

Amita Patnaik

Consulting or Advisory Role: Bayer AG, Novartis, Genentech (I), Roche (I), Merck
Research Funding: Merck (Inst), Pfizer (Inst), Eli Lilly (Inst), Plexikon (Inst), Corvus Pharmaceuticals (Inst), TESARO (Inst), AbbVie (Inst), Forty Seven (Inst), Five Prime Therapeutics (Inst), Infinity Pharmaceuticals (Inst), Proximagen (Inst), Pieris Pharmaceuticals (Inst), Surface Oncology (Inst), Livzon Pharmaceutical Group (Inst), Vigeo Therapeutics (Inst), Astellas Pharma (Inst), KLUS Pharma (Inst), Symphogen (Inst), Syndax (Inst)

Sumit A. Shah

Stock and Other Ownership Interests: Grand Rounds

Sreenivasa R. Chandana

Consulting or Advisory Role: Ipsen, AstraZeneca, Eli Lilly
Speakers' Bureau: Ipsen
Research Funding: Novocure (Inst)

Drew Rasco

Employment: START
Consulting or Advisory Role: TaiRx, Boehringer Ingelheim, Eli Lilly
Research Funding: Celgene (Inst), Millennium Pharmaceuticals (Inst), Rexahn Pharmaceuticals (Inst), Santa Maria Biotherapeutics (Inst), Five Prime Therapeutics (Inst), Pharmacyclics (Inst), Asana Biosciences (Inst), Eisai (Inst), Aeglea BioTherapeutics (Inst), Merck (Inst), Ascentage Pharma (Inst), Macrogenics (Inst), Apexian Pharmaceuticals (Inst), Birdie Pharmaceuticals (Inst), AbbVie (Inst), Constellation Pharmaceuticals (Inst), Syndax (Inst)
Travel, Accommodations, Expenses: Asana BioSciences

A. Dimitrios Colevas

Stock and Other Ownership Interests: Gilead Sciences, Pharmacyclics
Consulting or Advisory Role: COTA, KeyQuest Health, Loxo Oncology, Atara Biotherapeutics, Aduro Biotech, Cue Biopharma, Pfizer
Research Funding: Bristol-Myers Squibb, AstraZeneca, Innate Pharma, CellSight Technologies, Tessa Therapeutics

Timothy O'Rourke

Research Funding: Asana BioSciences (Inst)

Sujata Narayanan

Employment: Genentech
Stock and Other Ownership Interests: Roche

Kyriakos Papadopoulos

Consulting or Advisory Role: Bayer AG (Inst), ArQule (Inst)
Research Funding: AbbVie (Inst), MedImmune (Inst), Daiichi Sankyo (Inst), Regeneron Pharmaceuticals (Inst), Sanofi (Inst), ARMO BioSciences (Inst), ArQule (Inst), Calithera Biosciences (Inst), Curegenix (Inst), Incyte (Inst)Merck (Inst), Peloton Therapeutics (Inst), ADC Therapeutics (Inst), 3D Medicines (Inst), Formation Biologics (Inst), EMD Serono (Inst), Syros Pharmaceuticals (Inst), Mersana (Inst), OncoMed (Inst), MabSpace Biosciences (Inst), Jounce Therapeutics (Inst)

George A. Fisher

Stock and Other Ownership Interests: Seattle Genetics (I)
Honoraria: Merck, Genentech, Ipsen, Novartis
Consulting or Advisory Role: Ipsen, Genentech, Roche, Merck, Jounce Therapeutics
Research Funding: Newlink Genetics (Inst), XBiotech (Inst), Genentech (Inst), Roche (Inst), EpicentRx (Inst), Merck (Inst), Forty Seven (Inst), Aduro Biotech (Inst), Novartis (Inst)
Travel, Accommodations, Expenses: Merck Sharp & Dohme

Victor Villalobos

Consulting or Advisory Role: Janssen Pharmaceuticals, Eli Lilly, Novartis, AbbVie, Ignyta, Agios, Epizyme
Travel, Accommodations, Expenses: Eli Lilly, Janssen Pharmaceuticals, Xencor, GenMab, Apizyme

Susan S. Prohaska

Employment: Forty Seven
Stock and Other Ownership Interests: Forty Seven, Amgen
Consulting or Advisory Role: Forty Seven
Patents, Royalties, Other Intellectual Property: Inventor on patent developed at Stanford and assigned to Forty Seven
Other Relationship: Stanford University

Maureen Howard

Stock and Other Ownership Interests: Forty Seven
Consulting or Advisory Role: Cartherics
Patents, Royalties, Other Intellectual Property: Methods for Achieving Therapeutically Effective Doses of Anti-CD47 Agents, Stanford University, Nov 2014, WO 2014/1494772; Methods for Achieving Therapeutically Effective Doses of Anti-CD47 Agents, Stanford University, Feb 2014, US 502667359

Muralidhar Beeram

Honoraria: Genentech
Consulting or Advisory Role: Novartis (I)
Speakers' Bureau: Genentech
Research Funding: Eli Lilly (Inst), Zymeworks (Inst), Mersana Therapeutics (Inst)

Mark P. Chao

Employment: Forty Seven
Leadership: Forty Seven
Stock and Other Ownership Interests: Forty Seven, HepatX
Honoraria: D2G Oncology
Consulting or Advisory Role: Chimera Bioengineering
Patents, Royalties, Other Intellectual Property: Inventor on several patents from Stanford University licensed to Forty Seven, inventor on patents generated from Forty Seven
Travel, Accommodations, Expenses: Forty Seven

Balaji Agoram

Employment: Forty Seven
Stock and Other Ownership Interests: Forty Seven

James Y. Chen

Employment: Forty Seven
Stock and Other Ownership Interests: Forty Seven
Consulting or Advisory Role: Forty Seven
Research Funding: Forty Seven
Travel, Accommodations, Expenses: Forty Seven

Jie Huang

Employment: Forty Seven
Travel, Accommodations, Expenses: Forty Seven

Matthew Axt

Employment: Forty Seven
Employment: Kaiser Permanente (I)
Stock and Other Ownership Interests: Forty Seven

Jie Liu**Stock and Other Ownership Interests:** Forty Seven**Patents, Royalties, Other Intellectual Property:** Stanford/Forty Seven: CD47, SIRPa, CD99**Jens-Peter Volkmer****Employment:** Forty Seven**Stock and Other Ownership Interests:** Forty Seven**Patents, Royalties, Other Intellectual Property:** Co-inventor on technologies that have been licensed to Forty Seven**Ravindra Majeti****Employment:** Roche (I), Genentech (I)**Leadership:** Forty Seven**Stock and Other Ownership Interests:** Forty Seven, Roche (I), Genentech (I), Amgen (I), Imago BioSciences**Honoraria:** Roche, Genentech, Pharmacyclics**Consulting or Advisory Role:** Forty Seven, Biomarín**Research Funding:** Celgene, Bluestar Genomics**Patents, Royalties, Other Intellectual Property:** Numerous patents related to CD47 targeting and therapeutics that have been licensed to Forty Seven**Irving L. Weissman****Employment:** Forty Seven**Leadership:** Forty Seven**Stock and Other Ownership Interests:** Forty Seven**Consulting or Advisory Role:** Forty Seven**Patents, Royalties, Other Intellectual Property:** Many patents licensed by Stanford to Forty Seven, patents licensed to Systemix that were sold to Sandoz and Novartis, inventions for Systemix, patent pending in the brain stem cell field as part of Stem Cells cofounded in 1997 and sold in 2016**Travel, Accommodations, Expenses:** Forty Seven**Other Relationship:** Stem Cells (I)**Chris H. Takimoto****Employment:** Forty Seven**Leadership:** Forty Seven**Stock and Other Ownership Interests:** Johnson & Johnson, Forty Seven**Patents, Royalties, Other Intellectual Property:** Patent proposals filed related to using anti-CD47 antibodies as cancer therapy at Forty Seven**Travel, Accommodations, Expenses:** Forty Seven**Dana Supan****Research Funding:** Forty Seven (Inst)**Heather A. Wakelee****Honoraria:** Novartis, AstraZeneca**Research Funding:** Genentech (Inst), Roche (Inst), Pfizer (Inst), Eli Lilly (Inst), Celgene (Inst), AstraZeneca (Inst), MedImmune (Inst), Exelixis (Inst), Novartis (Inst), Clovis Oncology (Inst), Xcovery (Inst), Bristol-Myers Squibb (Inst), Gilead Sciences (Inst), Pharmacyclics (Inst), ACEA Biosciences (Inst)**Travel, Accommodations, Expenses:** AstraZeneca**Rhonda Aoki****Research Funding:** Forty Seven (Inst), Celldex Therapeutics (Inst)**Mark D. Pegram****Employment:** Loxo Oncology (I)**Consulting or Advisory Role:** Genentech, Pfizer, Novartis, Amgen, AstraZeneca, Merck**Travel, Accommodations, Expenses:** Genentech, Pfizer, Novartis, DAVA Pharmaceuticals**Sukhmani K. Padda****Consulting or Advisory Role:** Janssen Pharmaceuticals, G1 Therapeutics, AstraZeneca, AbbVie**Research Funding:** EpicentRx (Inst), Forty Seven (Inst), Bayer AG (Inst)

APPENDIX

Patient Eligibility

Additional patient eligibility criteria included having malignancy assessable for response by Response Evaluation Criteria in Solid Tumors (RECIST) for solid tumors or International Working Group criteria for lymphomas; ECOG performance status 0–2; life expectancy at least 3 months, and negative urine or serum pregnancy tests for all women of child-bearing potential. Exclusion criteria included primary brain tumors, patients with brain metastases, red blood cells (RBCs) transfusion dependence, hemoglobin < 9.5 g/dL, neutrophils < 1.0×10^9 /mL, AST/ALT > 5 times upper limits of normal (ULN), Bilirubin > 2.5 times ULN, serum creatinine > 1.5 times ULN, and significant medical conditions that would substantially increase the medical risks of participation in the study.

Methods

Dose-limiting toxicity exceptions: Grade 3 anemia; Grade 3 nausea, vomiting, diarrhea, or infusion reactions in the absence of premedication (including influenza-like symptoms, myalgias, fever, chills, headache, or acute pain) that resolves to \leq Grade 2 within \leq 72 hours after medical management (eg, supportive care) has been initiated; Grade 3 fatigue that resolves to \leq Grade 2 within 2 weeks on study; Grade 3 hyperbilirubinemia, isolated electrolyte abnormalities, or alanine aminotransferase and/or aspartate aminotransferase elevations that resolve to \leq Grade 2 with supportive care within 72 hours and is not associated with other clinically significant consequences; Grade 3 tumor lysis syndrome or related electrolyte disturbances (hyperkalemia, hypophosphatemia, hyperuricemia) that resolve to \leq Grade 2 within 7 days; Grade 3 or 4 lymphopenia.

Routine Laboratory Assessments

Laboratory assessments included routine blood counts with reticulocyte counts, general chemistry panels, serum uric acid, phosphorous, haptoglobin, D-Dimer, thrombin time, plasma fibrinogen, serum folic acid, prothrombin time, activated partial thromboplastin time, total iron and transferrin saturation, and urinalysis. All patients had a blood type and screen and direct antigen test performed at screening. Electrocardiograms were obtained at baseline and days 8, 29 and 57 of therapy. Peripheral blood smears were evaluated for blood cell morphology pre- and post-5F9 dosing on days 1 and 8 and

on days 2, 4, 8, 11, 15, 22 and 29. High resolution retinal photographs were performed at screening, and on days 1, 2, 4, 8, 9, 11 and 15 to assess for retinal microangiopathy; however, retinal imaging was halted in Part B due to a lack of correlative findings.

Pharmacokinetics and Anti-Drug Antibodies

Blood samples for pharmacokinetics (PK) monitoring were collected on day 1 pre-dose, and at the end of infusion and on day 8 at pre-dose and 0, 1, and 2 hours after the end of infusion. Additional PK samples were collected on days 9 and 30, and pre- and post-dose on days 11 (if dosed), 15, 22, 29, 36, 43, 50, and 57 and finally at the end of study treatment. Samples were shipped to a central laboratory for analysis of 5F9 concentration in serum using a validated ELISA Assay.¹⁵ The assay was specific to 5F9 and had a lower limit of quantification of approximately 200 ng/mL. Noncompartmental analysis (NCA) was performed using NCA for R software package to estimate PK parameters such as terminal half-life and area-under-the-concentration (AUC) curve.¹⁹

Anti-drug antibodies (ADA) were collected pre-dose and then every 4 weeks through week 17, and then on weeks 25, 33, 49 and 30 days after the last dose of 5F9. Samples were shipped to a central laboratory for ADA analysis.¹⁵ The ADA analysis followed a three-tiered approach, consisting of screening, confirmatory, and titer steps with pre-specified false positive rates of 5% and 1% for the screening and confirmatory steps, respectively. The cut points were generated using in-study samples.

Pharmacodynamics and Tumor Biopsies

Blood samples for CD47 receptor occupancy assessments on RBC and WBC were collected on day 1 and 8 pre- and post-dosing with 5F9 and on days 15, 22, 29, 36, 57 and end of study treatment. Receptor occupancy measurements were quantitated by using a flow cytometry assay with a tagged fluorescent anti-CD47 antibody to assess saturation of CD47 binding sites on RBC and WBC obtained from patients.

In the mandatory tumor biopsy cohort, patients were required to undergo mandatory tumor biopsies that were obtained pre-treatment and during week 6 of treatment. Tissue specimens were formalin-fixed and paraffin-embedded and subjected to immunohistochemical stains using an anti-IgG4 antibody. Because the levels of endogenous IgG4 levels are low in most tissues, treatment related changes could detect the presence of 5F9 in tissues during treatment.

TABLE A1. Summary of Geometric Mean (Geometric %CV) C_{max} and AUC Values after the First (Priming), Second, and Fifth Doses at All Dose Levels (Study SCI-CD47-001; Data Supplement)

Dose (mg/kg)	Week 1		Week 2			Week 5		
	n	C_{max} (mg/L)	n	AUC _{last} mg/L/d	C_{max} (mg/L)	n	AUC _{last} mg/L/d	C_{max} (mg/L)
0.1	1	NC	NA	NA	NA	NA	NA	NA
0.3	2	0.138 (NC)	1	0.00665 (NC)	0.170 (NC)	1	0.0246 (NC)	0.161 (NC)
1	6	0.682 (121)	6	0.141 (48.0)	5.83 (48.0)	6	2.89 (112)	10.6 (52.3)
3	6	0.637 (1,830)	3	33.7 (29.9)	45.3 (27.9)	2	80.0 (23.6)	58.7 (1.67)
10	3	0.573 (54.3)	3	577 (9.36)	177 (16.9)	3	1,560 (12.7)	368 (22.4)
20	35	0.758 (235)	35	898 (112)	411 (27.3)	33	2,560 (60.9)	728 (23.4)
30	3	0.776 (7.01)	2	724 (37.7)	586 (42.9)	2	5,080 (54.2)	1,090 (45.0)

Abbreviations: AUC, area-under-the-concentration; NA, not applicable; NC, not calculated